# PHARMACOKINETICS OF LEVOFLOXACIN IN COMPARISON TO THE RACEMIC MIXTURE OF OFLOXACIN IN MAN

M. Verho<sup>1</sup>, V. Malerczyk<sup>2</sup>, D. Damm<sup>3</sup> and K.-H. Lehr<sup>3</sup>

<sup>1</sup>Clinical Research, <sup>2</sup>Clinical Research/Biometrics, <sup>3</sup>Department of Pharma Research, 'Biochemistry, Hoechst AG, D-65926 Frankfurt, Germany

### SUMMARY

After oral administration of a single dose of 200 mg of levofloxacin and 400 mg racemic mixture of ofloxacin to 6 healthy male volunteers in a double-blind, randomised cross-over study, concentrations of the unchanged isomers were determined at various times in serum and urine, over 28 hours and 48 hours, respectively. Each dosing was followed by a wash-out period of one week. Ofloxacin concentrations were determined using an enantioselective and a non-enantioselective high pressure liquid chromatography (HPLC) assay. The two measurements obtained were compared by linear distribution independent regression, and were found to be equivalent.

Maximum serum concentration ( $C_{max}$ ) of levofloxacin after the administration of 200 mg of the levo-isomer was 2.42 mg/l (chiral derivatization HPLC, mean values); the corresponding area under the serum concentration-time curve (AUC<sub>0-28</sub>) was 17.0 mg x h/l. The corresponding  $C_{max}$  values after the administration of 400 mg ( $\pm$ )-isomer (chiral derivatization HPLC and reversed phased HPLC, mean values) were 2.05 mg/l, 1.98 mg/l and 4.41 mg/l for (-)-, (+)- and ( $\pm$ ) isomer, respectively. The AUCs<sub>0-28</sub> were 17.0, 14.6 and 32.7 mg x h/l, respectively. The pharmacokinetics of the (-)- and (+)-isomer were shown to be almost equal. In serum and urine no reracemisation of the

Author for correspondence:

M. Verho Hoechst Aktiengesellschaft Clinical Research, H 840 D-65926 Frankfurt am Main Germany (-)-isomer to a racemic mixture was observed. General tolerability was good; no side effects were reported.

## **KEY WORDS**

levofloxacin, ofloxacin, racemic mixture, pharmacokinetics

# INTRODUCTION

Drug enantiomers can have different pharmacokinetic /1,2/ and pharmacodynamic /3,4/ properties.

Ofloxacin, a gyrase inhibitor with an extremely broad antibacterial spectrum /5-7/, has a chiral C-atom at the C3-position and therefore consists of two optically active isomers /8/. The *in vitro* antibacterial activities of the (-)-form are 8 to 128 times that of (+)-ofloxacin and about twice that of the racemic mixture of ofloxacin against grampositive and gram-negative bacteria tested /8,9/. The (-)-form is highly active against isolates of *E. coli, Citrobacter freundii, K. pneumoniae, Enterobacter cloacae, H. influenzae* and *B. catarrhalis* with MIC<sub>90</sub>s ranging from 0.05 to 0.39 mg/l. The (+)-isomer is poorly active against *Staphylococcus* spp., *Streptococcus* spp. and *Enterococcus* spp. with MIC<sub>90</sub>s ranging from 50 - >100 mg/l. It is active against gram-negative bacterial isolates but the activity is at least 30 times less than that of the (-)-isomer /10/.

The aim of the present study was to investigate the pharmaco-kinetics of the pure (-)-form<sup>1</sup> in comparison to the racemic mixture, in order to detect possible differences in the pharmacokinetic characteristics. A second goal was to investigate whether an *in vivo* reracemisation from the (-)-form to the racemic mixture takes place in the body.

# SUBJECTS AND METHODS

Six healthy male volunteers entered the study. Their mean age was  $41 \pm 8$  years. They had normal body build with a mean weight of  $77 \pm 7$  kg and a mean height of  $174 \pm 5$  cm. All volunteers gave their

<sup>&</sup>lt;sup>1</sup> 0.3% of the (+)-form was detected as contamination in this form (Lehr, personal communication).

consent in writing after being informed by the physician of the nature, purpose and possible risks of the trial. All volunteers underwent an initial physical examination including standard safety checks that was repeated 24 hours after the last application; there were no pathological findings.

The volunteers received a single oral dose of 200 mg levofloxacin or 400 mg of the racemic mixture in a double-blind, randomised cross-over study design. Each dosing was followed by a wash-out period of one week before the next dosing.

Blood samples were taken from an antecubital vein before dosing and at the following times after dosing: 5 min, 15 min, 30 min, 45 min, 1 h, 1.25 h, 1.5 h, 1.75 h, 2 h, 2.5 h, 3 h, 3.5 h, 4 h, 5 h, 6 h, 7 h, 9 h, 12 h, 14 h, 24 h, 25 h and 28 hours. 7 ml blood (2 aliquots of 1.5 ml of serum) was taken at each sampling time. Urine was collected in 4-hourly fractions for 12 hours, thereafter in 12-hourly fractions for a further 36 hours.

Two HPLC-methods were applied to this study. (-)-Ofloxacin (levofloxacin) and (+)-ofloxacin concentrations in serum and urine were determined using high pressure liquid chromatography (HPLC) after chiral derivatization /11/. In addition, (±)-ofloxacin concentrations were determined using a non-enantioselective HPLC-method /12/.

The imprecision between day of the enantioselective assay determined in the concentration range from 15 ng to 5000 ng per ml serum was between 5.3 and 12.6% for (-)-ofloxacin and between 4.9 and 12.9% for (+)-ofloxacin. In urine, the corresponding values were between 2.9 and 18.8% for (-)-ofloxacin and between 2.9 and 16.1% for (+)-ofloxacin determined in the concentration range from 0.15 to 50 mg/l. The detection limits for (-)- and (+)-ofloxacin in serum were 10 ng/ml and in urine 50 ng/ml and 70 ng/ml, respectively /11/.

The imprecision between day of the non-enantioselective assay determined in the concentration range from 10 ng to 2000 ng per ml serum was between 1.9 and 10.2% for  $(\pm)$ -ofloxacin. In urine, the corresponding values were between 2.1 and 11.1% for  $(\pm)$ -ofloxacin determined in the concentration range from 0.5 to 500 mg/l. The detection limit for  $(\pm)$ -ofloxacin was 4 ng/ml in serum and 200 ng/ml in urine.

The concentration-time profiles of ofloxacin were best described by a two-compartment open model /13/, which was fitted to the data by means of a computer programme for non-linear approximations /14/.

# **RESULTS**

# Method comparisons

Measurement comparisons using linear distribution independent regression analysis were calculated for serum and urine according to Passing and Bablok /15/, shown in Figure 1 for serum.

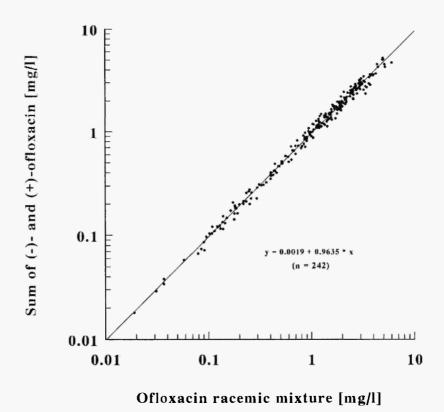


Fig. 1: Measurement comparison for serum between the original HPLC method for the racemic mixture and the sum of the two enantiomers determined by means of the enantiospecific HPLC method.

The sums of (-)- and (+)-forms in both serum and urine were compared directly with the data from original HPLC concentrations. An excellent correspondence (r = 0.986 for serum, r = 0.994 for urine; p < 0.001 for both) between HPLC methods was found; both regression lines are not statistically different from the unity line (slope 1, intercept 0).

# **Pharmacokinetics**

The time courses of ofloxacin concentrations in serum after both dosings are shown in Figures 2 and 3. The corresponding pharmacokinetic data are given in Table 1.

The terminal half-life, AUC values and the urinary recovery of the (+)-isomer were slightly less than the corresponding values of the (-)-form in the 400 mg racemate tablet. The pharmacokinetic variables of 200 mg levofloxacin and 400 mg tablet estimated as (-)-isomer showed no differences at all.

Neither in urine nor serum was there any evidence of reracemisation of the (-)-isomer to the corresponding (+)-form.

# Tolerability

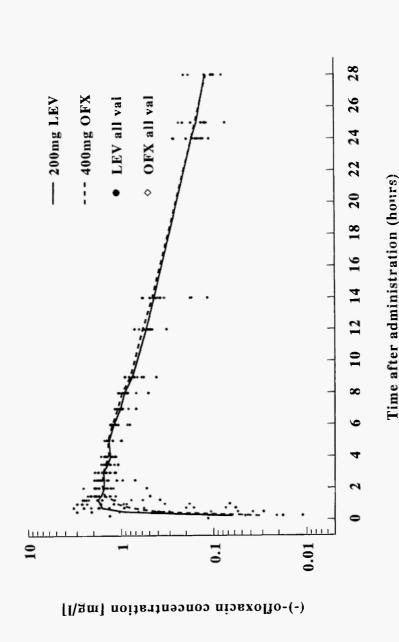
Adverse events did not occur in any subject during the trial period; no clinically relevant changes were noted in haematological, biochemical or urine analyses.

### DISCUSSION

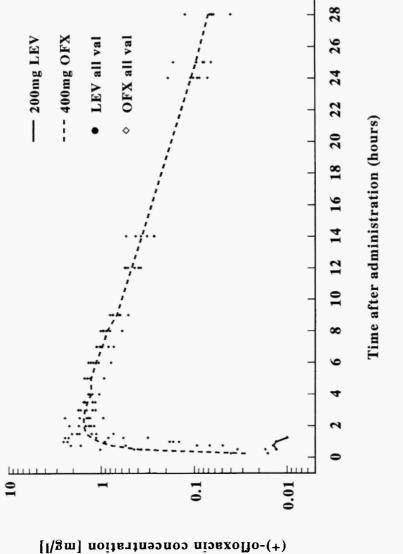
Levofloxacin, the S-(-)-isomer of ofloxacin, has an antibacterial activity twice as high /8,9/ and an anti-DNA activity twice as high than ofloxacin, the racemic mixture /16/. In the treatment of complicated urinary tract infections levofloxacin in a dose of 100 mg t.i.d. proved as effective as ofloxacin 200 mg t.i.d. /17/.

The enatioselective disposition of ofloxacin after administration of the racemic mixture has been investigated in animals and man. Okazaki et al. /18/ investigated the serum concentration profiles in rat, monkey and dog. They found that in rat the (R)-ofloxacin predominates, whereas the opposite is the case in monkeys. In dogs no differences in the pharmacokinetics of the enationers were observed.

The enatioselective disposition of ofloxacin was investigated in healthy subjects after oral administration of 200 mg (±)-ofloxacin /19/.



Mean serum concentration curves of the two ofloxacin formulations administered (200 mg levofloxacin, 400 mg racemic mixture): a) analysed as (-)-isomer by chinal derivatization HPLC; b) analyted as (+)-isomer by chinal danvaization HPLC; c) analysed by reverse 1 phase HPLC. Fig. 2:



Mean serum concentration curves of the (-)- and (+)-isomer analysed from the 400 mg racemic mixture.

Pharmacokinetic data of (-)- and (+)-ofloxacin and the racemic mixture after oral administration of 200 mg

| Variables                           |                  | 200 mg levoflox acin       |                  | 94                         | 400 mg racemic mixture |                  |
|-------------------------------------|------------------|----------------------------|------------------|----------------------------|------------------------|------------------|
|                                     | chiral derivati  | chiral derivatization HPLC | rewrsed phase    | chiral derivatization HPLC | zation HPLC            | reversed phase   |
|                                     | m.oj-(-)         | (+)-form <sup>2</sup>      |                  | (-)-form                   | m:oj-(+)               |                  |
| C <sub>max</sub> [m,3/1]            | 2.42 ± 0.54      | 0.01                       | 2.50 ± 0.42      | 2 05 ± 0.56                | 1.98 ± 0.53            | 4.41 ± 1.09      |
| C <sub>12</sub> [mg/.]              | $0.54 \pm 0.10$  |                            | $0.54 \pm 0.07$  | $0.59 \pm 0.10$            | $0.47 \pm 0.08$        | $1.06 \pm 0.17$  |
| C <sub>24</sub> [mg/ <sub>1</sub> ] | $0.17 \pm 0.05$  |                            | $0.17 \pm 0.04$  | $0.17 \pm 0.06$            | $0.11 \pm 0.04$        | $0.30 \pm 0.10$  |
| tmax [h]                            | $1.4 \pm 0.7$    | 8.0                        | $1.4 \pm 0.7$    | $1.8 \pm 0.7$              | $1.8 \pm 0.7$          | $1.8 \pm 0.7$    |
| 1½ ß [h]                            | $6.5 \pm 0.9$    |                            | $6.8 \pm 0.6$    | $6.5 \pm 1.4$              | $5.6 \pm 1.2$          | $6.0 \pm 0.9$    |
| AUC <sub>0-28</sub> [mg*h/l]        | $17.0 \pm 3.2$   | 0.01                       | $17.1 \pm 2.1$   | $17.0 \pm 2.6$             | $14.6 \pm 2.3$         | $32.7 \pm 4.5$   |
|                                     |                  |                            |                  | 31.6:                      | $31.6 \pm 3.5$         |                  |
| AUDC [mg*h/l]                       | 17.9 ± 3.6       |                            | $18.1 \pm 2.4$   | $17.8 \pm 3.0$             | $14.8 \pm 2.6$         | $34.1 \pm 4.7$   |
|                                     |                  |                            |                  | 32.7                       | $32.7 \pm 40$          |                  |
| CLto!/f[ml/min]                     | $192.3 \pm 36.3$ |                            | $186.7 \pm 23.7$ | $191.4 \pm 32.8$           | $230.4 \pm 39.0$       | 198.5 ± 25.1     |
| URINARY RECOVERY                    |                  |                            |                  |                            |                        |                  |
| gm                                  | $151.3 \pm 16.8$ | 29±60                      | $182.1 \pm 21.5$ | $153.4 \pm 18.9$           | $144.4 \pm 180$        | $337.6 \pm 37.9$ |
| % of dose                           | 75.7             |                            | 91.4             | 38.4                       | 36.1                   | 84.4             |
|                                     |                  |                            |                  | 74.5                       | 5                      |                  |

See footnote 1

No differences in the peak concentration or time to reach peak concentration were observed, suggesting that the absorption of ofloxacin is not stereoselective in humans; however renal clearance was different. Renal clearance of the R-(+)-isomer was significantly greater than that of the S-(-)-isomer. The half-life of the S-(-)-isomer was statistically significantly longer than that of the R-(+)-isomer.

The results of this study indicate that in the racemic mixture the (+)-isomer possesses a slight trend to a decreased half-life, a slight reduction of the AUC-value and a slight decrease in urinary recovery of the unchanged drug. These differences, however, are minimal and without any relevance with respect to clinical application. The half-lives and urinary recoveries of both formulations are in the same range as reported previously /20-23/.

No differences were seen in the levofloxacin pharmacokinetic variables after the administration of 200 mg of the pure (-)-isomer or 400 mg of the racemic mixture (See Table 1) and no reracemisation occurred in the (-)-isomer to the racemic mixture as shown in the urinary excretion of the drug.

The good antibacterial activity of levofloxacin in comparison to the racemic mixture makes this substance a very interesting new development. However its clinical superiority as compared to the racemate has still to be confirmed in the clinical setting.

# REFERENCES

- Eichelbaum M, Mikus G, Vogelgesang B. Pharmacokinetics of (+), (-), and (±) verapamil after intravenous administration. Br J Clin Pharmacol 1984; 17: 453-458.
- Wingard LB Jr, O'Reilly RA, Levy G. Pharmacokinetics of warfarin enantiomers: A search for intrasubject correlations. Clin Pharmacol Ther 1978; 23: 212-217.
- 3. Ho IK, Harris RA. Mechanism of action of barbiturates. Ann Rev Pharmacol Toxicol 1981; 21: 83-111.
- 4. Cotzias GC, Papavasiliou PS, Gellene R. Modification of parkinsonism chronic treatment with L-dopa. N Engl J Med 1969; 280: 337-345.
- Cullmann W, Stieglitz M, Baars B, Opferkuch W. Comparative evaluation of recently developed quinolone compounds - with a note on the frequency of resistant mutants. Chemotherapy 1985; 31: 19-28.
- 6. Goossens H, De Mol P, Coignau H, Levy J, Grados O, Ghysels G, Innocent H, Butzler JP. Comparative in vitro activities of aztreonam, ciprofloxacin, norfloxacin, ofloxacin, HR 810 (a new cephalosporin), RU 28965 (a new

- macrolide), and other agents against enteropathogens. Antimicrob Agents Chemother 1985; 27: 388-392.
- Seibert G, Limbert M, Klesel N. Comparison of the antibacterial in vitro and in vivo activity of Ofloxacin (HOE 280 DL 8280) and Nalidixic acid analogues. Eur J Clin Microbiol 1983; 2: 548-553.
- Une T, Fujimoto T, Sato K, Osada Y. In vitro activity of DR-3355, an optically active ofloxacin. Antimicrob Agents Chemother 1988; 32: 1336-1340.
- Hayakawa I, Atarashi S, Yokohama S, Imamura M, Sakano KI, Furukawa M. Synthesis and antibacterial activities of optically active ofloxacin. Antimicrob Agents Chemother 1986; 29: 163-164.
- Tanaka M, Otsuki M, Une T, Nishino T. In-vitro and in-vivo activity of DR-3355, an optically active isomer of ofloxacin. J Antimicrob Chemother 1990; 26: 659-666.
- Lehr KH, Damm P. Quantification of the enantiomers of ofloxacin in biological fluids by high-performance liquid chromatography. J Chromatogr 1988: 425: 153-161.
- Lameire N, Rosenkranz B, Malerczyk V, Lehr KH, Veys N, Ringoir S. Ofloxacin pharmacokinetics in chronic renal failure and dialysis. Clin Pharmacokinet 1991; 21: 357-371.
- Wagner JG. Fundamentals of Clinical Pharmacokinetics. 2nd Ed. Hamilton, Illinois: Drug Intelligence Publications, 1979.
- 14. Von Hattingberg HM, Brockmeier D, Kreuter G. A rotating iterative procedure (RIP) for estimating hybrid constants in multi-compartment analysis on desk computers. Eur J Clin Pharmacol 1977; 11: 381-388.
- Passing H, Bablok W. A new biometrical procedure for testing the equality of measurements from two different analytical methods. Application of linear regression procedures for method comparison studies in clinical chemistry, Part I. J Clin Chem Clin Biochem 1983; 21: 709-720.
- Imamura M, Shibamura S, Hayakawa I, Osada Y. Inhibition of DNA gyrase by optically active ofloxacin. Antimicrob Agents Chemother 1987; 31: 325-327.
- 17. Kawada Y, Kumamoto Y, Aso Y, Machida T, Saito I, Kawamura N, Ohkoshi M, Naide Y, Kawabe K, Hisazumi H, Okada K, Kamidono S, Ohmori H, Usui T, Kagawa S, Fujita Y, Kumazawa J, Ohi Y, Ueno K, Ogawa N. Comparative study on levofloxacin and ofloxacin in complicated urinary tract infections. Chemotherapy (Tokyo) 1992; 40 (Suppl 3): 230-248.
- Okazaki O, Kurata T, Hakusui H, Tachizawa H. Species-related stereoselective disposition of ofloxacin in the rat, dog and monkey. Xenobiotica 1992; 22: 439-450.
- Okazaki O, Kojima C, Hakusui H, Nakashima M. Enantioselective disposition of ofloxacin in humans. Antimicrob Agents Chemother 1991; 35: 2106-2109.
- Dagrosa E, Seeger K, Malerczyk V, Lameire N. Pharmakokinetik von Ofloxacin (HOE 280). FAC-Fortschr Antimikr Antineoplast Chemother 1984; 3-5: 665-671.

- 21. Verho M, Malerczyk V, Dagrosa E, Korn A. Dose linearity and other pharmacokinetics of ofloxacin: a new, broad-spectrum antimicrobial agent. Pharmatherapeutica 1985; 4: 376-382.
- 22. Verho M, Malerczyk V, Dagrosa E, Korn A. The effect of food on the pharmacokinetics of ofloxacin. Curr Med Res Opin 1986; 10: 166-171.
- 23. Dagrosa EE, Verho M, Malerczyk V, De Looze S, Hajdu P, Toyodera K. Multiple-dose pharmacokinetics of ofloxacin, a new broad-spectrum antimicrobial agent. Clin Ther 1986; 8: 632-645.